

Integrative and Comparative Biology

Integrative and Comparative Biology, volume 61, number 2, pp. 521–537 doi:10.1093/icb/icab082

Society for Integrative and Comparative Biology

SYMPOSIUM

Future Tail Tales: A Forward-Looking, Integrative Perspective on Tail Research

M. J. Schwaner ¹* S. T. Hsieh,[†] I. Braasch,[‡] S. Bradley,[§] C. B. Campos,[¶] C. E. Collins,[¶] C. M. Donatelli,[∥] F. E. Fish,^{|||} O. E. Fitch,[‡] B. E. Flammang ⁽¹⁾,^{**} B. E. Jackson,^{††} A. Jusufi,^{‡‡} P. J. Mekdara,^{§§} A. Patel,^{¶¶} B. J. Swalla ⁽¹⁾,^{|||||} M. Vickaryous ⁽¹⁾ and C. P. McGowan^{***}

*Department of Ecology and Evolutionary Biology, University of California Irvine, Irvine, CA 92697, USA; [†]Department of Biology, Temple University, Philadelphia, PA 19122, USA; [†]Department of Integrative Biology and Program in Ecology, Evolution, and Behavior (EEB), Michigan State University, East Lansing, MI 48824, USA; [§]Department of Biomedical Science, University of Guelph, Guelph N1G 2W1, Canada; ⁹Department of Biological Sciences, Sacramento State University, Sacramento, CA 95819, USA; ^{||}Department of Biology, University of Ottawa, Ontario K1N 6N5, Canada; ^{|||}Department of Biology, West Chester University, West Chester, PA 19383, USA; **Department of Biological Sciences, New Jersey Institute of Technology, Newark, NJ 07102, USA; ^{††}Department of Biological and Environmental Sciences, Longwood University, Farmville, VA 23909, USA; ^{‡‡}Max Planck Institute for Intelligent Systems, Stuttgart 70569, Germany; ^{§§}National Institute of Neurological Disorders and Stroke, National Institutes of Health, Bethesda, MD 20892, USA; ¹⁵Department of Electrical Engineering, University of Cape Town, Cape Town 7701, South Africa; ^{||||||}Department of Biology, University of Washington, Seattle, WA 98195, USA; ***Department of Integrative Anatomical Sciences, University of Southern California, Los Angeles, CA 90033, USA

From the symposium "An evolutionary tail: Evo-Devo, structure, and function of post-anal appendages" presented at the virtual annual meeting of the Society for Integrative and Comparative Biology, January 3–7, 2020.

¹E-mail: mjschwan@uci.edu

Synopsis Synopsis Tails are a defining characteristic of chordates and show enormous diversity in function and shape. Although chordate tails share a common evolutionary and genetic-developmental origin, tails are extremely versatile in morphology and function. For example, tails can be short or long, thin or thick, and feathered or spiked, and they can be used for propulsion, communication, or balancing, and they mediate in predator–prey outcomes. Depending on the species of animal the tail is attached to, it can have extraordinarily multi-functional purposes. Despite its morphological diversity and broad functional roles, tails have not received similar scientific attention as, for example, the paired appendages such as legs or fins. This forward-looking review article is a first step toward interdisciplinary scientific synthesis in tail research. We discuss the importance of tail research in relation to five topics: (1) evolution and development, (2) regeneration, (3) functional morphology, (4) sensorimotor control, and (5) computational and physical models. Within each of these areas, we highlight areas of research and combinations of long-standing and new experimental approaches to move the field of tail research forward. To best advance a holistic understanding of tail evolution and function, it is imperative to embrace an interdisciplinary approach, re-integrating traditionally siloed fields around discussions on tail-related research.

Introduction

Although the post-anal tail is considered a defining chordate characteristic, tails are also present in taxa outside of this phylum. Tails have extraordinarily diverse functions, including, but not limited to use as a fifth limb, as a visual signal for warning and courtship, and as an essential physiological and morphological driver for functions related to propulsion, stability and maneuverability, prehension, energy storage, and thermoregulation (Wake and Dresner 1967; Aleksiuk 1970; Lindsey 1978; Fish 1979, 1982; Hickman 1979; Tucker 1992; Thomas 1997; Patel et

Advance Access publication May 17, 2021

[©] The Author(s) 2021. Published by Oxford University Press on behalf of the Society for Integrative and Comparative Biology.

All rights reserved. For permissions please email: journals.permissions@oup.com.

al. 2016). In other words, the tail plays multiple roles critical to many animals' survival. Such multifunctionality of the tail has captured the imagination of engineers and roboticists, leading to remarkable advances in the development of versatile bioinspired robotic systems (Kopman et al. 2015; Patel and Boje 2015; Saab et al. 2018; Zhu et al. 2019; Fish 2020).

Despite its apparent importance, tails are vastly understudied. For example, we still have little understanding for why tails are conserved, reduced, or lost in certain lineages; or why analogous structures exist among invertebrates. Likewise, developmental pathways and regenerative mechanisms of tails are only recently gaining greater attention. Within biomechanics research, the tail is often ignored, with a greater focus toward paired appendage function. Yet, tails precede the evolution of paired appendages by ~200 million years (Donoghue and Keating 2014). As such, increasing our understanding of tail function, development, and evolution may provide valuable insight into the physiological costs and benefits of also using paired appendages, as well as how the holistic coordination of multiple appendage types-a concept common to fish swimming research but not elsewhere-enables smooth motion. Research on tails continues in a broad range of fields, from Evolutionary Developmental Biology (EvoDevo) to behavior to robotics, but these fields remain largely isolated from one another; yet there is immense conceptual overlap among these fields that have transformative potential if collaboration was more common. It is therefore imperative that we reintegrate these fields to fully capture the complexity and diversity of tails from an interdisciplinary viewpoint.

With this forward-looking perspective paper, we aim to cross boundaries between research fields, summarize what is known, and highlight exciting future tail research directions with the hope of inspiring greater interdisciplinary discourse. Here we outline five key areas to deepen and broaden our knowledge on the breath of tail and tail-related topics. We discuss the current state of the field including knowledge gaps, potential areas for future studies, and interdisciplinary research approaches in (1) evolution and development, (2) regeneration, (3) functional morphology, (4) sensorimotor control, and (5) computational and physical models.

Evolution and development

Although the notochord is a homologous structure across chordates and an integral part of post-anal

tails, questions remain about the homology of skeletal tail structures. A clear definition of what comprises the tail is surprisingly elusive. If we define the tail as the "post-anal extension of the body axis," that creates a predicament since that definition in fishes would include parts of the dorsal fin and the anal and caudal fin as part of the tail. Developmental processes may provide some clarity, as there is evidence pointing toward unique genetic modules that individually control the development of these unpaired fins (Letelier et al. 2018). Thus, we should not lump all fins together as part of the tail. Sallan (2016) proposes that the fish caudal region is actually a combination of two components: (1) the caudal fin and (2) the "tail," defined as the postvertebral notochord extension. This theory further postulates that differential outgrowth of these two structures generated the variation in the caudal region across fishes, including tetrapods (Sallan 2016). Moving forward, a careful examination of the gene regulatory networks (GRNs) controlling the development of these individual components of the postanal caudal region will thus be necessary to evaluate homologies among chordate and vertebrate tails.

Compared with caudal structures, paired appendages that led to the origin of tetrapods have received a lot more attention in EvoDevo research. When anatomy alone was not enough to determine homology between the structure of the paired pectoral and pelvic fins in fishes and tetrapod fore and hindlimbs, development of molecular techniques allowed deep genomic homologies to be drawn between paired fins and limbs (Shubin et al. 1997; Mercader 2007; Yano and Tamura 2013; Gehrke and Shubin 2016). Similar use of genetic techniques could provide missing links in determining homologies among chordate tail structures. Analyzing developmental GRNs in the evolutionarily old caudal region may also shed light on the evolutionary origin of the younger paired appendages that first emerged in an ancestor of jawed vertebrates, potentially by co-opting genetic programs from other body appendages such as tails.

There are three key GRN components known to be involved in tail development across chordates: *Homeobox (Hox)* and *T-box* transcription factor genes, as well as Hedgehog signaling genes. However, large gaps still exist in our understanding of tail-specific regulatory landscapes. Here, we highlight what is already known about expression and control of these genes in relation to tail Evo-Devo and point out what is yet to be understood.

Hox genes in most vertebrates are organized into four duplicated (paralogous) clusters (Hox A, B, C, and D). Together, these pattern the antero-posterior body plan and the proximo-distal axis of paired appendages in the developing embryo (reviewed in, e.g., Burke et al. 1995; Duboule 1998; Holland 2013). Gene order along the chromosome reflects expression order along the body axis; the first Hox genes are expressed in the anterior of the embryo, and the last Hox genes are expressed in the most posterior regions of the embryo as well as the most distal ends of the fin/limb bud. In paired fins/limbs, only Hox A and Hox D cluster genes are expressed in limbs in vertebrates (Freitas et al. 2012; Tulenko et al. 2016; Longellato et al. 2018; Desanlis et al. 2020). We still lack an understanding of how the posterior Hox genes are expressed and regulated during caudal development across vertebrates. It has become clear that posterior genes of paralogy groups Hox 9-13 are expressed in post-anal tails of all chordates as well as in hemichordates (Fodor et al. 2021). Most of our current knowledge comes from mouse models, which suggest a major role for posterior Hox B and Hox C cluster genes in tail development (Economides et al. 2003; Aires et al. 2019).

The key notochord gene brachyury (bra), generally known as Tbxt, is essential for chordate tail development. This is referred to by many names: bra in tunicates, T in mouse, Xbra in frog, and no tail (*ntl*) in fishes. This gene was present as one copy in early chordates. Two paralogous copies (Tbxta, Tbxtb) appeared during the early two rounds of vertebrate genome duplication, before one of these two paralogs (Tbxta) was secondarily lost again in the tetrapod common ancestor (Amemiya et al. 2013). The expression pattern of *Tbxt* genes in the notochord and the tail bud is conserved across chordates (Hermann 1995; Corbo et al. 1997; Thisse 2001). Loss of function of Tbxt genes leads to complete loss of tail in zebrafish and incomplete tail development in mice (Wilson et al. 1993; Schulte-Merker 1995). In zebrafish tail development, tbxta and tbxtb are required for expression of Wnt signaling genes during posterior mesoderm formation (Martin and Kimelman 2008). A recent study shows that although bra has been considered a master regulator of notochord development, bra alone is not sufficient to regulate notochord development in Ciona, highlighting the importance of further research into understanding the chordate tail GRN (Reeves et al. 2021).

In the Hedgehog signaling pathway, the vertebrate ligand gene *sonic hedgehog* (*shh*) plays many roles during development, one of which is establishing the Zone of Polarizing Activity (ZPA) that patterns fin and limb development (Gehrke and Shubin 2016). Its expression in the paired fin/limb is regulated by the ZPA regulatory sequence enhancer and in fish, a similar enhancer has been found to specify shh expression during the development of the unpaired dorsal fin (Letelier et al. 2018). The vital role of shh in paired and unpaired appendage development in fishes suggests that it could also play an important, yet elusive, role in fish tail development (Hadzhiev et al. 2007). Hedgehog signaling has been implicated in tail development in zebrafish (Hadzhiev et al. 2007), and loss of shh signaling in mouse models leads to abnormal tails (El Shahawy et al. 2019), but the specific involvement of this pathway in vertebrate caudal development has yet to be determined. Hedgehog signaling does not seem to be involved in tail development in ascidians, but does play a role in amphioxus. More work is thus necessary to understand the evolution of Hedgehog signaling in chordate caudal development (Di Gregorio 2020).

There is considerable overlap in gene expression in the tail and the paired appendages. However, this overlap is not necessarily an indication of a cooption, perhaps it is an indication of common processes (e.g., the similarity between posterior and distal axis extension). Despite the obvious similarities in gene expression, such as posterior Hox gene expression, few studies have focused on the possible caudal origin hypothesis: that the paired appendages are a co-option of GRNs specifying the posterior body axis (Shubin et al. 1997). There are several other competing hypotheses that the paired appendages and their GRNs originated from (1) the gills, (2) a hypothetical lateral fin fold, or (3) the median unpaired fins (dorsal and anal fin). No single hypothesis stands out because there are data supporting all of them (Abe and Ota 2017; Letelier et al. 2018) and some have hypothesized a combination of these origins (Diogo Sleight and Gillis 2020). 2020; Understanding the genes that pattern the nonvertebrate chordate tail compared with those that pattern the vertebrate tail as well as the vertebrate fin/limb will elucidate whether the tail really is the "first fin" and help answer a longstanding question that has interested scientists for over a century.

Although the post-anal tail is a key characteristic of chordates, the tail has also been reduced or secondarily lost in many lineages, as exemplified in birds and in the absence of a great ape tail. It is possible that a thorough investigation of instances of tail loss across chordate evolution would help us understand human tail loss, as well as other cases of tail reduction or loss among vertebrates (e.g., in frogs and birds). Different from ancestral, multisegmented vertebrate tails, birds have fused caudal vertebrae that form pygostyles, which support tail feathers important for maneuvering during flight. Development of these pygostyles only happens after hatching (Rashid et al. 2018), indicating that birds retain the ancestral tail morphology in-ovo. In all cases of vertebrate tail loss, the tail is present throughout embryonic/larval development, but then is lost during development into the adult form-as can be seen among frogs and humans. Extending to non-vertebrate chordates, tunicates generally follow this pattern as well: the embryonic tail is lost upon reaching adulthood. However, there are some ascidian tunicates that have also lost the embryonic tail. Understanding the evolutionary and developmental processes that have led to instances of tail loss or tail vertebrae fusion across chordates could lead to a comprehensive study that encompasses the comparison of tail loss across many examples in chordate evolution.

Regeneration

Among some vertebrates, the tail demonstrates an unusual capacity for wound repair and regeneration (Bellairs and Bryant 1985; Higham et al. 2013; Jacyniak et al. 2017; Gordeev et al. 2020; Verissimo et al. 2020; Xu et al. 2020). Instead of scarring, these species are able to spontaneously regrow a replacement. Regeneration begins as an aggregation of proliferating cells called a blastema. Blastema cells eventually give rise to most new tail tissues. Although blastema-mediated regeneration is also associated with paired appendage regrowth among some anamniotes, the tail is the only appendage capable of regenerating in amniotes. Further, unlike the limbs, the tail (in various species of plethodontid salamanders and lepidosaurs) can be self-amputated or autotomized-an anti-predation mechanism that permits the controlled release of a portion of the tail. Once released, the detached tail will thrash, thereby providing a distraction as the prey escapes. It is worth noting, however, that while tail autotomy is often paired with tail regeneration, the two mechanisms are independent of one-another. More specifically, while some species capable of tail autotomy are incapable of tail regeneration (e.g., some snakes and amphisbaenians), other species that do not autotomize are capable of tail regeneration (e.g., teleost fish, non-plethodontid- and some plethodontidsalamanders, tadpoles, and crocodylians).

While tail regeneration recreates the overall shape and function of the original appendage, the fidelity of the replacement often varies. Whereas zebrafish and salamanders effectively replicate the pattern and structure of the amputated tail, reptiles do not. For example, leopard geckos primarily regenerate autotomized tails through cartilage and fat deposits, retaining a key function for fat storage, and are morphologically simple relative to their original counterparts (Gilbert et al. 2013). Among lizards, the loss in fidelity is most readily observed in the spinal cord (regenerated spinal cords lack gray matter), skeleton (bony vertebrae are replaced by a hollow cone of cartilage), skin (variation in the pattern of scalation), and a simplified musculoskeletal arrangement. The regenerated tails of *Sphenodon* and crocodylians are even less exact replacements, composed largely of a cartilaginous cone surrounded by connective tissue with limited skeletal muscle.

The amount of physiological energy and resources that are diverted into regenerating a lost tail is often indicative of their importance. Organisms that rely on tails for functional roles such as locomotion, antipredation, and fat storage are predicted to suffer reduced fitness following autotomy (Barr et al. 2019; Triay-Portella et al. 2019). As such, energy that is allotted to maintaining regular body functioning may be funneled into caudal tissue regeneration, sometimes through multiple tail regeneration events in a lifetime (each loss occurs progressively closer to the tail base), or even "re-regeneration" of the exact same tail region (Barr et al. 2019). Energy allocation strategies between tail regeneration and reproduction vary among species and even between sex within a species (Dial and Fitzpatrick 1981; Salvador et al. 1995; Fox et al. 1998; Yurewicz and Wilbur 2004; Triay-Portella et al. 2019). Alternatively, birds use tail feather autotomy and regeneration as antipredator strategy, which is found to be physiologically inexpensive and does not affect flight performance (Møller et al. 2006; Johansson and Hedenstrom 2009). These suggest an evolutionary cost-benefit approach to the facilitation of tail regeneration, and a hierarchy of energy allocation according to different pressures, potentially explaining why some species may not be capable of tail regeneration, or "choose" to suppress this capability entirely, in favor of other biological investments. However, this hypothesis remains to be tested.

Our understanding of the genetic and molecular mechanisms of tail regeneration is still limited and deeper analysis among different organisms is required to fully understand tail regeneration over various taxa (vertebrates, chordates, planarians, etc.). So far, studies examining regeneration in planarians, lizards, and tadpoles show very different pathways to regeneration: increased expression of genes that control cell proliferation, critical roles for transforming growth factor β signaling pathways, upregulation of Wnt signals, and less expression of inflammatory-immune genes (Ho and Whitman 2008; Almuedo-Castillo et al. 2012; Hutchins et al. 2014; Vitulo et al. 2016; Xu et al. 2020). A substantial amount of work has been done with plethodontid salamanders to understand the factors that influence tail autotomy and regeneration (Marvin 2010; Marvin and Lewis 2013). These findings lead to a need of further investigation of similarity in these pathways and genes in organisms capable of successful regeneration across the phylogenetic tree. Technology in genomics and molecular science can be useful tools to compare and contrast the mechanisms and gene expressions leading to observed variations in tail regeneration efficiency. For example, it allows for comparison of the restorative regeneration of planarians, that rebuild their complete lost anatomy, with modified anatomy in regenerated lizard tails (Elliot and Alvarado 2012). Moreover, in order to further investigate the depth of such questions there must be a greater effort to genome sequence species aside from currently used models such as Anolis carolinensis (has a complete and annotated genome sequence) and Eublepharis macularius (Alföldi et al. 2011; Xiong et al. 2016). Thus far, it is unknown how new structures can proliferate, differentiate, and organize while surrounded by adult cells. Comparative studies among species with high and low regenerative fidelity could elucidate why some adult animals can regenerate entire structures and why some lose this capacity.

Tail regeneration is hypothesized to have broad consequences for ecology and behavior; however, direct experimental evidence for these consequences is limited. Much of what we know is based on locomotor performance of salamanders and a small number of lizard species, namely leopard geckos and green anoles (e.g., Higham et al. 2013; Hsieh 2016). The results of these studies provide little consensus. Salamanders, for example, regenerate tails with fidelity; can regain full swimming capacities with only 50% regeneration; and regeneration speed is dependent on temperature, body size, and amount of tail length lost (Marvin 2010, 2011, 2013; Marvin and Lewis 2013; Joven et al. 2019). In contrast, most lizards lose locomotor capacity following autotomy, and it is slowly regained during regeneration. This directly impacts predator avoidance and mate selection during and even after regeneration (Gillis et al. 2009; Higham et al. 2013; Jagnandan et al. 2014; Hsieh 2016; Jagnandan and Higham 2017). Equivocal evidence from tadpoles, however, indicates that tail damage and regeneration may or may not 525

have functional consequences on adult body size and adult locomotor performance (Ding et al. 2014; Koch and Wilcoxen 2019; Zamora-Camacho et al. 2019), emphasizing the need for comparative approaches to understanding the interplay between tail regeneration and ecology.

To rectify the gap in current knowledge, tailregenerating species across a phylogenetic tree should be included in a cross-disciplinary analysis. For example, arboreal lizards that use tails to help climb and balance and lizards that use tails to enhance sprint speed or communicate would be better models to measure how regenerated structures support movement. Given the variation in the capacity to (1) autotomize using fracture planes, (2) the fidelity of regenerated structures, and (3) the secondarily simplified regenerated tails of lizards, we suggest leveraging the diversity of lizard habitat niches to measure the value of tails (Pianka 1973; Skeels et al. 2020). If a comparative and ecological approach is taken to sample the diversity of tail regeneration capacity in vertebrates, then the evolutionary pressures of maintaining or evolving the ability to regenerate a lost appendage could be better known.

Functional morphology

The tail provides a variety of functions that attest to its evolutionary and ecological importance. As noted earlier in this review, while some organisms-including humans—have reduced or lost the tail, a large percentage of vertebrates have maintained, elongated, elaborated, or repurposed this caudal extremity. In early vertebrates that descended from their chordate ancestors, the tail initially functioned as a device for swimming and stabilizing the body from both internal and external perturbations (Webb 2002). Embellishment of the tail with a broad caudal fin enhanced the propulsive effect by its interaction with the fluid environment (Flammang 2014). The hydrodynamics of the undulatory tail motions with a substantial caudal fin resulted in greater accelerations, swimming velocities, and efficiency due to action on an increased mass of fluid, facilitating momentum transfer from the axial musculature and vertebral column to the tail, to the water (Drucker and Jensen 1996; Müller et al. 2001; Tytell and Lauder 2004). Additionally, some fish can move outside of the water with the help of their tail (Swanson and Gibb 2004; Hsieh 2010; Gibb et al. 2013; Ashley-Ross et al. 2014), whereas certain salamanders can perform tail-assisted jumps (Hessell and Nishikawa 2017; Brown and Deban 2020).

The importance of a tail and its terminal fin for high performance swimming is a quintessential example of morphological and physiological modifications by evolution to a common successful form (Hildebrand 1995; Liem et al. 2001; Kardong 2019). The examples of the shark, ichthyosaur, and dolphin illustrate the effectiveness of this adaptation via convergent evolution. These disparate organisms exhibit similarities in shape and mechanics of the tail, despite their phylogenetic separation. This textbook example demonstrates how similar functional requirements of the tail are met by different clades that have evolved as identical solutions to analogous environmental challenges. For example, tail morphology is associated with environmental pressures selecting on the need to operate in the open ocean for fast, efficient propulsion. Convergence in morphology is then also exploited for similar trophic opportunities by highly derived aquatic predators.

As vertebrates moved out the water and conquered the terrestrial environment with limbed locomotion, the tail evolved new functions rather than simply being dragged on the ground (McInroe et al. 2016). Fast running animals use the tail as an aerodynamic inertial appendage to rapidly maneuver when changing direction (Wilson et al. 2013; Patel and Braae 2014). Saltatorial animals and bipedal runners employ the tail as a counterbalance (Alexander and Vernon 1975; Gillis et al. 2013; O'Connor et al. 2014). Even slow-moving tetrapods make use of the tail as an effective passive defensive strategy by sacrificing the extremity to allow the body an opportunity for escape (Vitt et al. 1977; Humphreys and Ruxton 2019). Rapid whip-like motions of the tail can provide a weapon for hunting and defense, which can be further enhanced by the incorporation of a massive tail club as displayed by the extinct Ankylosaurus (Arbour 2009; Oliver et al. 2013). A highly mobile tail affords the ability for prehension. The seahorse (Hippocampus) can wrap the tail around stationary objects for anchorage (Hale 1996; Neutens et al. 2014). In addition, a prehensile tail facilitates climbing in a variety of tetrapods except for birds (Hickman 1979; Lemelin 1995; Luger et al. 2020). However, the reduced fused pygostyle of birds supports the tail feathers that are necessary to foster stability in flight and swimming (Gatesy and Dial 1996; Felice and O'Connor 2014). Finally, the tail with its high surface-to-volume ratio can act as a thermal window in liberating excess heat, particularly from mammals (Fish 1979; Hickman 1979).

Two extant tetrapod vertebrate lineages that evolved flight, birds and bats, further modified tails

for aerodynamic and other functions. Reduced fused pygostyle of birds supports the tail feathers while allowing unrivaled levels of modification of the full tail shape and position (Gatesy and Dial 1996) allowing it to variably act as an added wing, rudder, stabilizer, and drag reducer (e.g., Maybury et al. 2001; Maybury and Rayner 2001; Sachs 2007; Usherwood et al. 2020). Bat tails, made of a membrane stretched between hind limbs and including the caudal vertebrae, also provide a flight control surface (Gardiner et al. 2011). In both cases, the added lift and reduced drag provided by the tails decrease energetic costs relative to wing-only flight. Bird tails can also provide visual and even auditory signals for courtship and predator avoidance (Woodland et al. 1980; Clark et al. 2011) and can aid prey capture (e.g., Jackson and Elgar 1993). Bat tail membranes can even act as an insect net (Webster and Griffin 1962).

For so long, anatomical descriptions and proposed functions of the tail were investigated by simple dissections and morphometric measurements. There have been considerable advances in medical imaging and computational and physical modeling techniques that have opened a new window into understanding the relationship between form and function of the caudal extremity. computed tomography (CT) scans now allow a non-invasive means of examining both the external and internal composition of hard and soft anatomical features of tails (Watkins-Colwell et al. 2018; Buser et al. 2020). These scans can be used to create 3D models of skeletal anatomy which can be used in a variety of ways. Finite element analysis can then take these 3D models and apply loads to different bones and joints to investigate strain propagation and range of motion (Hsieh et al. 2005). 3D geometric morphometrics and other statistical techniques can tell us how morphology changes within a group or over evolutionary time (Buser et al. 2018). The powerful technique of X-ray of moving morphology can be directed toward analysis of the complex arrangement of the multiple joints and their movements by the tail (Brainerd et al. 2010). Investigations of structural mechanics (i.e., stress and strain) of real specimens can assess tail strength, flexibility, and range of motion and provide the data to examine tail autotomy and regeneration (Hsieh et al. 2005; Peixoto et al. 2019). Models generated from CT scans can be 3D printed and tested on universal testing machines to investigate how shape restricts movement (Connors et al. 2019). The importance of the tail as a thermal window can be investigated with thermographic cameras (McCafferty 2007).

With incorporation of new technologies and a perspective that views tails as an important feature

in mechanics and physiology of organisms, there are opportunities to address long-standing questions and develop new possibilities for research. Tails function as a transmission directing the forces generated from muscles to the eventual exchange of momentum from swimming animals to the fluid environment. How this is accomplished has not been fully understood. Likewise, the role of the tail in stability and maneuverability is incomplete and needs to be appropriately modeled. Evolutionarily, what anatomical and morphological adaptations lead to the diversity of tail functions? Can we use the wide range of morphological data from CT scans along with kinematic data and computational modeling to create a system that "predicts" function in extant and extinct animals? Such questions can direct research projects along several avenues that demonstrate how the tail is an integral component of animals and essential in their biology.

Sensorimotor control

The wide variety of behaviors for which animals use their tails, as highlighted in the sections above, require them to extract sensory information from the surrounding environment, filter and sort through that information to determine the environmental relevance, and adapt its movements to reach a predetermined course. During locomotion, the behavior is quite complex because all of these steps have to be achieved while the animal is moving, thus the animal is constantly adjusting its motor output to changing visual, olfactory, auditory, and mechanosensory feedback (*Fig. 1*; Goulding 2009; Huston and Jayaraman 2011; Wyart and Knafo 2015; Koch et al. 2018).

The sensorimotor system lies in the ability to transform multiple sensory modalities into a locomotor movement (Schouenborg and Weng 1994; Levinsson et al. 1999; Nishikawa et al. 2007; Schouenborg 2008; Goulding 2009). Multisensory processing is essential to adapting to noisy sensory environments, enhances the robustness of the animal's motor output, and increases efficiency of sensorimotor tasks (Wyart and Knafo 2015). Studying multisensory integration with respect to the animal's tail can provide insight in how animals use this appendage to navigate and overcome obstacles in the real world. An example of multisensory processing through sensorimotor integration, with an essential role of the tail, is the startle escape response seen in fish, which is an integral behavior needed for the fish's survival (Hale et al. 2002). The precise timing and movements of a fish's body and tail into a cshape, coupled with a tail flick, to generate a fast escape response after detection of a predator is driven directly by the sensorimotor system (Hale et al. 2002; Tytell and Lauder 2002; Faber et al. 2006; McHenry et al. 2009). This behavioral response consists of coordination of neural activity, rhythmic pattern generating networks, contraction of muscles, movement of the fish, and feedback networks to modulate the speed and direction of the movement. Yet, the contribution of the multisensory information and the neural circuits that regulate the behavioral output is still not completely understood (Hale et al. 2002; Nishikawa et al. 2007; Schuster 2012).

Although tail related motor tasks are key components of locomotion in many animals (e.g., Lauder 2000; Higham and Russell 2010; Freymiller et al. 2019), developing practical experimental models that require one to reproduce behaviorally relevant multisensory environments with consistent motor outputs is strenuous (Goulding 2009; Wyart and Knafo 2015). On the one hand, some of the reasons behind the lack of tail sensorimotor control studies are that tail morphology and tail-related behavioral repertoires exhibit variable and complex patterns of motor activity, which make practical considerations of tail-related experimental designs quite difficult to develop. On the other hand, the neural circuits that drive these tail movements have been less tractable due to the complexity of those circuits with highly variable patterns of activity and unreliable experimental tools to identify them. In addition, analyzing complex datasets with many levels of kinematic parameters per animal, including interactions between animals, raises an important technical challenge (Wyart and Knafo 2015; Mathis et al. 2018). One solution proposed in the field of neuromechanics is to use computational and physical models, such as bioinspired robotics (e.g., Jusufi et al. 2010; Libby et al. 2012; Patel et al. 2016; Zhu et al. 2019; Lin et al. 2021), to help predict some of the behavioral computations without observing the variability of non-linear motor outputs from the animal's tail. However, sensory integration and the neural circuits that underlie the movements of the tail are an essential component of sensorimotor studies, which cannot fully be explained by simplified computational and physical models.

Reflex-based tail movements are well suited for experimental analysis and can provide insights into how a moderately complex sensorimotor system can generate tail-based locomotion. At this scale, sensorimotor integration at the "high-level" cortical areas, normally associated with many complex networks of neuronal projections and interactions in the central nervous system, can be simplified to "low-level"

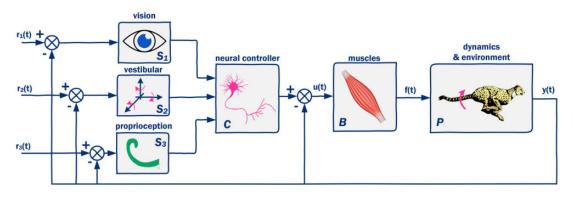


Fig. 1. Neuromechanics of tail responses in animal locomotion. Diagram encompassing underlying neuromechanical control. The tail behavior of a model animal (e.g., cheetah) expressed through a feedback control theoretic framework. The cheetah body interacts with the environment (P) for locomotion. Three outer control loops are proposed, the first is the visual processing of the cheetah chasing the prey (S1), the second is the vestibular processing of the cheetah trying to maintain its balance (S2) and the third is proprioceptive processing of the cheetah tail (S3) using muscle spindles. These parallel sensor mappings are processed by a neural controller (C) which then activates the tail musculature (B) to produce forces on the cheetah body. These muscles are also activated by high frequency mechanical feedback when interacting with the environment.

regions such as the spinal cord. Sherrington's pioneering studies have provided insights into the nature of such neural pathways, with illumination of the reflex arc and the control of reciprocal motor actions through inhibitory neural networks in vertebrates (Sherrington 1908; Brown and Sherrington 1912; Sherrington 1913). In addition to early studies from Eccles, Lundburg, Brown, and Jankowska (Brown and Sherrington 1912; Brown 1914; Lundberg 1979; Jankowska 2001), there has been increasing evidence to suggest that sensorimotor integration also occurs at the spinal cord (Grillner et al. 1991; Schouenborg et al. 1995; Mentel et al. 2006; Schouenborg 2008; Nakanishi and Whelan 2012; Hilde et al. 2016; Uemura et al. 2020; Picton et al. 2021), a region that has been originally hypothesized to contain intrinsic networks of neurons that generate rhythmic locomotor-like patterns of activity known as the central pattern generator (CPG) (Brown and Sherrington 1912; Brown 1914; Lundberg 1979; Jankowska 2001). Sensory inputs can modulate and shape the outputs of the CPG. However, whether the CPG is involved in driving tail movements needs further exploration. Studying sensorimotor control of tails, even at this reasonably low level, still requires one to select a behaviorally relevant multisensory environment that is tail-driven and can be reproducible consistently.

In practice, an important goal of neurobiologists is to understand how the nervous system is organized and functions to generate the locomotor movement (Hale et al. 2002; Nishikawa et al. 2007; Goulding 2009). Therefore, we propose some key questions to understanding the sensorimotor control of tails: (1) How do neural circuits that contain millions of neurons integrate multisensory information and flexibly contribute to specific motor patterns involved in tail-driven locomotion? (2) How are these ensembles of neurons organized and coordinated to interact with rhythm and pattern generating circuits to produce the overall motor output or behavior? (3) What is the neural computation of the circuit underlying tail-driven movements and how are these movements encoded? (4) What is the behavioral computation that includes both the active and passive mechanical properties of the movement? And (5) are these sensorimotor systems evolutionarily tuned to the locomotor mechanics and are they conserved across taxa?

Sensorimotor integration is highly dynamic where the animal is constantly responding to changes in the environment and updating its sensory inputs according to its behavioral output: for example, as a kangaroo rat detects a snake strike, vestibular and visual stimuli are constantly changing in the middle of an escape jump (Freymiller et al. 2019). Ideally, one would be able to capture every level of behavior with real-time monitoring of the sensorimotor integration as it occurs in the circuits, but practical considerations of such an experiment often make it challenging and difficult. In classical experimental settings, to probe these neuronal networks responsible for generating some of the behavioral output, the animal must often be restrained, paralyzed, or reduced to a "fictive" experimental preparation, often without sensory inputs (e.g., semi-intact, decerebrated, deafferented, and ablation), to allow recordings of active neurons—a combination of electrophysiology, pharmacological manipulation, and anatomical identification is often required

(Nishikawa et al. 2007; Goulding 2009; Wyart and Knafo 2015; Koch et al. 2018). Even in reduced animal preparations, these experimental designs are limited by difficulties in manipulating the neurons directly in the circuits, overcoming the large locomotor-driven activity of the CPG, and producing reproducible behaviors consistently. In recent years, the convergence of developmental genetics and physiological and behavioral systems approaches has expanded the possibilities of directly mapping behavioral sensorimotor computations onto specific neural circuits, targeting circuits of interests genetically, and monitoring the activity in vivo as the sensorimotor integration occurs (Goulding 2009; Wyart and Knafo 2015; Dobrott et al. 2019). As we move forward in the field, sophisticated genetic manipulations in powerful model systems such as mice and zebrafish have allowed for more precise characterization of the neural populations and its function in behavior. Tail sensorimotor control can essentially be dissected and pieced together using modern tools such as ontogenetics, chemogenetics, optogenetics, multi-array electrophysiology, and automated behavioral tracking systems that utilize deep learning artificial neural networks (Levine et al. 2014; Mathis et al. 2018; Dobrott et al. 2019; Lotfollahi et al. 2020).

Computational and physical models

In contrast to querying what animals are doing to implement use of the tail in a complex system, an alternative approach applies computational and physical modeling approaches. The advantage of these methods is that they permit simplification of the remainder of the system to a degree unachievable in biological systems, to identify governing principles driving tail function. In other words, they promote a template-based approach that permits progressive increases in complexity of the examined system toward the biological anchor (Full and Koditschek 1999). When combined with experimental robotics, this can provide a powerful framework for achieving valuable biological insight and facilitating discovery of novel physics principles (Flammang and Porter 2011; Aguilar et al. 2016) and a better understanding of sensorimotor control of tail movements (Fig. 1).

A classic example of a templates-anchor approach is that of the locomotor spring-mass model used for running gaits. The spring-mass model holds no clear representation to the multi-segmented, multi-material, highly complicated organism; but by virtue of its simplification to a mere mass mounted on a spring, it is capable of capturing a surprising sophistication of dynamic behaviors inherent to legged locomotor systems (e.g., Blickhan 1989; McMahon and Cheng 1990; Schmitt and Holmes 2000; Gever et al. 2002). By gradually adding layers of complexity to this template, it then became possible to identify the roles of other subtleties critical to legged locomotion, that otherwise would have been impossible to isolate, such as leg retraction kinematics, hip torque, and stiffness modulation (e.g., Seyfarth et al. 2003; Daley and Biewener 2006; Spence et al. 2010; Shen and Seipel 2012). Like the spring-mass model, a tail template model of comparable simplicity could provide insight into basic physical principles guiding its function. For example, whereas a simple mass on a stiff or spring-tether may be sufficient to capture its dynamics for basic inertial function, imbuing greater sophistication via a distributed mass-spring model can reveal how subtle changes in position or curvature impact function. These, and increasingly more complex tail models, could then advance our understanding of tail mechanics and functional evolution.

Somewhat more sophisticated template-based models can be tested using physical models. For example, these physical models can be obtained using a combination of micro-CT scans and 3D printing, to derive shapes of varying levels of sophistication and with different stiffness and elasticity (Esposito et al. 2012; Porter et al. 2015; Flammang et al. 2017; Behm et al. 2018). In doing so, it becomes possible to isolate aspects of a tail's complex morphology and material properties and examine how each component contributes to the overall function of the entire structure. Likewise, it allows testing of biologically "impossible" shapes, to query why they might not exist. Porter et al. (2015) relied on this approach to discover why seahorse tails are square, and also why most other vertebrates have tails with a round cross-sectional geometry. Another important application of physical models is that they can be used to "revive" extinct species (e.g., McInroe et al. 2016; Johnson and Carter 2019; Ibrahim et al. 2020). Using extant species as a guide, it becomes possible to reconstruct aspects of an extinct animal's ecology and evolutionary history with a greater degree of specificity than previously possible.

Implementation of these models digitally in a computationally simulated environment provides additional insight on appendage–environmental interactions. Although this approach is already commonly used to quantify tail function during flight and swimming (Borazjani and Sotiropoulos 2008, 2009) using computational fluid dynamics models, it is much less frequently applied during explorations of terrestrial tail use. Implementation of finite element methods embedded within a discrete element model would permit simultaneous quantification of appendage force production and environment response-a topic that has been gaining increasing attention only during the last decade or so. Additionally, the digital realm provides an opportunity for rapid computational evolution of appendage morphology (Moore et al. 2013), and thus an ability to test a diverse range of shapes subjected to different kinematics and/or environmental parameters that would otherwise be extremely time-consuming if working exclusively with physical models. Everincreasing computational power and availability of open-source modeling software (e.g., Large-scale Atomic/Molecular Massively Parallel Simulator [LAMMPS] and LAMMPS Improved for General Granular and Granular Heat Transfer Simulations [LIGGGHTS]; Plimpton 1995; Kloss et al. 2012) is making this approach ever more tractable.

The field of bioinspired and biomimetic robotics was born from the recognition of the incredible versatility, stability, and grace with which animals move through the world, and the subsequent desire to emulate their form and function in a wide variety of models and robots. Because of the broad range of tasks for which animals use their tails, researchers have focused on this appendage on several robotic platforms (e.g., Chang-Sui et al. 2011; Lauder et al. 2011; Zhang et al. 2016; Rosic et al. 2017; Saab et al. 2018; Liu and Ben-Tzvi 2021). Terrestrial bipedal and quadrupedal robots and vehicles have been equipped with tails for manipulation, climbing, maneuvering, and balance (Spenko et al. 2008; Chang-Sui et al. 2011; Rone and Ben-Tzvi 2015; Zhang et al. 2016; Flammang et al. 2017; Saab et al. 2018). And there has been increasing interest in design and development of aquatic robotics and vehicles that use fishlike, tail-propelled swimming (e.g., Triantafyllou and Triantafyllou 1995; Long et al. 2006; Lauder et al. 2011; Kopman et al. 2015; Rosic et al. 2017; Lin et al. 2021), motivated by a need for robots that are capable of prolonged, efficient swimming, and/or making difficult and smooth maneuvers. As a whole, experimental robotics is largely driven by task-based goals: using the addition of a physical tail to add greater functionality for manipulation or balance, or to allow the robot to make swifter turns or maneuver mid-air (Chang-Sui et al. 2011; Briggs et al. 2012; Libby et al. 2012; Rone and Ben-Tzvi 2014). The next-generation tail models should include more than rigid body dynamics alone, and instead

integrate multi-joint flexibility and stiffness properties that afford animal tails their adaptability and 3D dexterity. An area of priority is the development of actuators that more closely model muscle mechanics. These actuators could then be tested in future tail models, reducing weight and increasing smoothness of motion. With these considerations in mind, the implementation of tail models using soft robotics could be another productive direction to pursue. Soft actuators allow for shape changes and bending actuation with potentially infinite degrees of freedom, thus facilitating modeling of prehensile tails, while soft sensors enable sensory feedback to provide insight on the neuromechanics of locomotion, such as the closed loop control of body caudal fin swimming of soft robotic fish (Lin et al. 2021), in addition to permitting measurement of curvature in conditions where videography is not feasible (e.g., nocturnal and low visibility medium).

In conjunction with bioinspired robotics is an emerging field of robophysics (Aguilar et al. 2016). A robophysical approach is highly complementary to experimental robotics, but instead of being driven by optimization of task-driven goals, it seeks to use instances of success and failure to reveal underlying principles of the interaction. A broad exploration of parameter space also permits the discovery of new locomotor strategies as well as novel physics principles that might otherwise be impossible to deduce when working with the full complexity of a system (e.g., Li et al. 2009; Marvi et al. 2014; Aguilar et al. 2016). Where robophysics is most powerful when dissecting contributions from multiple, complex sources is of interest. This includes understanding the role of the tail relative to the applied dynamics of other appendages on the body, or even adding in considerations of the complex, natural environment in which the animal is performing a behavior of interest, which may shift unexpectedly as the animal moves (e.g., Schwaner et al. 2021, this issue; Shield et al. 2021, this issue).

Conclusion

Tails are a common characteristic in animals but the diversity in development and repertoire of functional behaviors varies greatly across and within taxa. Depending on the species, the tail can have a singular or multi-functional purpose. In addition, tails exhibit a wide range of dexterity and morphological characteristics. The development, evolution, shape, and function of an appendage that seems otherwise relatively simple, such as a tail, has led researchers, scientists, and engineers to a proliferation of bio-inspired designs of dynamic robotic devices that use tails for an even wider range of tasks. Yet, there are many questions to be answered to understand the evolution, development, sensorimotor capabilities, morphology, ecology, and functions of tails across the phylogenetic tree and applications in bioinspired designs.

Expanding our knowledge of tails has important implications and has the potential to answer many long-standing questions. For example, understanding the morphology and genetic foundation of post-anal appendages can elucidate the evolutionary origin(s) and functions of tails, and opens opportunities to begin to grasp concepts like secondary loss, as seen in humans. In addition, understanding the relationship between the form and function of the tail, we can make informed predictions of tail function based on their appearance, with implications for studies of both extant and extinct species. Kinematics, sensorimotor, and morphological tail characteristics can also be used to further develop computational models and bioinspired robotics. Lastly, comparison of next generation tail models with vertebral column models may have important medical implications for treatments for human vertebral dysfunction (e.g., Ishihara 1996; Handa et al. 1997; Iatridis et al. 1999; Demers et al. 2004; Han et al. 2008).

Tail research crosses many disciplines, and here we presented just a focused review of some of them. We discussed the current state and future directions for tail-related research in five key topic areas. Although they were presented as distinct categories, these topics are inherently interrelated. For example, the post-anal tail is a phylum-defining component of the chordate bauplan, and similar genetic pathways are found to be important for both tail development and regeneration (Martin and Kimelman 2008; Vitulo et al. 2016). Tail regeneration-or lack thereof-has important consequences for sensorimotor control strategies and functional morphology, which in turn inform experimental robotics and robophysics inquiry. Examining tail function using a combination of these approaches, as well as computational and physical modeling, will provide deep insight into the evolutionary pathways and functional importance of tails to the locomotor system and beyond. As a result, a re-integration of biological disciplines is critical to the progress of multiple fields. This can be facilitated by future symposia, workshops, and seminars, focused on advancements of tail and tail-related research. Such interactions among tail researchers can serve as a foundation for cross-disciplinary research collaborations to shed light on the form, function, EvoDevo, and technical applications of animal tails.

Authors' contribution

S.T.H., C.P.M., and M.J.S.: initial manuscript ideas. S.T.H., C.P.M., M.J.S., S.B., C.B.C., C.E.C., C.M.D., F.E.F., O.E.F., B.E.F., B.E.J., A.J., P.J.M., A.P., B.J.S., and M.V.: conceptualizing ideas. S.T.H., C.P.M., M.J.S., I.B., S.B., C.B.C., C.E.C., C.M.D., F.E.F., O.E.F., B.E.F., B.E.J., A.J., P.J.M., A.P., B.J.S., and M.V.: first manuscript draft (I.B., O.E.F., and B.J.S.: evolution and development. S.B., C.B.C., C.E.C., and M.V.: regeneration. C.M.D., F.E.F., and B.E.J.: functional morphology. B.E.F., A.J., P.J.M., and A.P.: sensorimotor control. S.T.H., C.P.M., and M.J.S.: computational and physical modeling). S.T.H., C.P.M., M.J.S., I.B., S.B., C.B.C., C.E.C., C.M.D., F.E.F., O.E.F., B.E.F., B.E.J., A.J., P.J.M., A.P., B.J.S., and M.V.: editing manuscript. S.T.H., C.P.M., and M.J.S.: final edits.

Acknowledgments

This forward-looking review was a product of a workshop series in conjunction with a symposium, entitled "An Evolutionary Tail: EvoDevo, Structure, and Function of Post-Anal Appendages" as part of the Society for Integrative and Comparative Biology Virtual Annual Meeting, held January and February 2021. The authors would like to thank all participants of the first workshop whose discussions were helpful with composing this manuscript (in alphabetical order): Pasha van Bijlert, Luke Bollinger, Bob Cieri, Jaden Clark, Renee Dickie, Gabrielle Russo, Johanna Schultz, Shaylee Smith, Marina Vollin, Jesse Young, and Vanessa Young.

Funding

This work was supported by the National Science Foundation (NSF), grant number #2029523 [to C.P.M. and S.T.H.]. In addition, work was supported by NSF #1453106 [to S.T.H.] and #1553550 [to C.P.M.].

Conflict of interest

Authors declare no conflict of interest.

References

- Abe G, Ota KG. 2017. Evolutionary developmental transition from median to paired morphology of vertebrate fins: perspectives from twin-tail goldfish. Dev Biol 427:251–7.
- Aguilar J, Zhang T, Qian F, Kingsbury M, McInroe B, Mazouchova N, Li C, Maladen R, Gong C, Travers M, et al. 2016. A review on locomotion robophysics: the study of movement at the intersection of robotics, soft matter and dynamical systems. Rep Prog Phys 79:110001.

- Aires R, de Lemos L, Nóvoa A, Jurberg AD, Mascrez B, Duboule D, Mallo M. 2019. Tail bud progenitor activity relies on a network comprising Gdf11, Lin28, and Hox13 genes. Dev Cell 48:383–95.
- Aleksiuk M. 1970. The function of the tail as a fat storage depot in the beaver (Castor canadensis). J Mammal 51:145–8.
- Alexander RM, Vernon A. 1975. The mechanics of hopping by kangaroos (Macropodidae). J Zool 177:265–303.
- Alföldi J, Di Palma F, Grabherr M. 2011. The genome of the green anole lizard and a comparative analysis with birds and mammals. Nature 477:587–91.
- Almuedo-Castillo M, Sureda-Gomez M, Adell T. 2012. Wnt signaling in planarians: new answers to old questions. Int J Dev Biol 56:53–65.
- Amemiya CT, Alföldi J, Lee AP, Fan S, Philippe H, MacCallum I, Braasch I, Manousaki T, Schneider I, Rohner N, et al. 2013. The African coelacanth genome provides insights into tetrapod evolution. Nature 496:311–6.
- Arbour VM. 2009. Estimating impact forces of tail club strikes by ankylosaurid dinosaurs. PLoS ONE 4:e6738.
- Ashley-Ross MA, Perlman BM, Gibb AC, Long JH. 2014. Jumping sans legs: does elastic energy storage by the vertebral column power terrestrial jumps in bony fishes? Zoology 117:7–18.
- Barr JI, Boisvert CA, Somaweera R, Trinajstic K, Bateman PW. 2019. Re-regeneration to reduce negative effects associated with tail loss in lizards. Sci Rep 9:18717.
- Behm JE, Waite BR, Hsieh ST, Helmus MR. 2018. Benefits and limitations of three-dimensional printing technology for ecological research. BMC Ecol 18:32.
- Bellairs ADA, Bryant SV. 1985. Autotomy and regeneration in reptiles. In: Gans, CBillett, F, editors. Biology of the reptilia. New York (NY): Wiley. p. 301–410.
- Blickhan R. 1989. The spring-mass model for running and hopping. J Biomech 22:1217–27.
- Borazjani I, Sotiropoulos F. 2008. Numerical investigation of the hydrodynamics of carangiform swimming in the transitional and inertial flow regimes. J Exp Biol 211:1541–58.
- Borazjani I, Sotiropoulos F. 2009. Numerical investigation of the hydrodynamics of anguilliform swimming in the transitional and inertial flow regimes. J Exp Biol 212:576–92.
- Brainerd EL, Baier DB, Gatesy SM, Hedrick TL, Metzger KA, Gilbert SL, Crisco JJ. 2010. X-ray reconstruction of moving morphology (XROMM): precision, accuracy and applications in comparative biomechanics research. J Exp Zool A Ecol Genet Physiol 313:262–79.
- Briggs RJ, Lee M, Haberland M, Kim S. 2012. Tail in biomimetics design: analysis, simulation, and experiment. IEEE/ RSJ International Conference on Intelligent Robots and Systems, Vilamoura, Algarve, Portugal.
- Brown TG. 1914. On the nature of the fundamental activity of the nervous centres. J Physiol 48:18–46.
- Brown CE, Deban SM. 2020. Jumping in arboreal salamanders: a possible tradeoff between takeoff velocity and in-air posture. Zoology 138:125724.
- Brown TG, Sherrington CS. 1912. The rule of reflex response in the limb reflexes of the mammal and its exceptions. J Physiol 44:125–30.

- Burke AC, Nelson CE, Morgan BA, Tabin C. 1995. Hox genes and the evolution of vertebrate axial morphology. Development 121:333–46.
- Buser TJ, Boyd OF, Cortés Á, Donatelli CM, Kolmann MA, Luparell JL, Pfeiffenberger JA, Sidlauskas BL, Summers AP. 2020. The natural historian's guide to the CT galaxy: stepby-step instructions for preparing and analyzing computed tomographic (CT) data using cross-platform, open access software. Integr Organ Biol 2:obaa009.
- Buser TJ, Sidlauskas BL, Summers AP. 2018. 2D or not 2D? Testing the utility of 2D vs. 3D landmark data in geometric morphometrics of the sculpin subfamily Oligocottinae (Pisces; Cottoidea). Anat Rec 301:806–18.
- Chang-Sui E, Libby T, Tomizuka M, Full RJ. 2011. A lizardinspired active tail enables rapid manuevers and dynamic stabilization in a terrestrial robot. IEEE/RSJ International Conference on Intelligent Robots and Systems, San Francisco, CA.
- Clark CJ, Elias DO, Prum RO. 2011. Aeroelastic flutter produces hummingbird feather songs. Science 333:1430–3.
- Connors M, Yang T, Hosny A, Deng Z, Yazdandoost F, Massaadi H, Eernisse D, Mirzaeifar R, Dean MN, Weaver JC, et al. 2019. Bioinspired design of flexible armor based on chiton scales. Nat Commun 10:1–13.
- Corbo JC, Levine M, Zeller RW. 1997. Characterization of a notochord-specific enhancer from the Brachyury promoter region of the ascidian, *Ciona intestinalis*. Development 124:589–602.
- Daley MA, Biewener AA. 2006. Running over rough terrain reveals limb control for intrinsic stability. Proc Natl Acad Sci USA 103:15681–6.
- Demers CN, Antoniou J, Mwale F. 2004. Value and limitations of using the bovine tail as a model for the human lumbar spine. Spine 29:2793–9.
- Desanlis I, Kherdjemil Y, Mayran A, Boukouch Y, Gentile C, Sheth R, Zeller R, Drouin J, Kmita M. 2020. HOX13-dependent chromatin accessibility underlies the transition towards the digit development program. Nat Commun 11:2491.
- Di Gregorio A. 2020. Chapter Eleven—The notochord gene regulatory network in chordate evolution: conservation and divergence from *Ciona* to vertebrates. Curr Top Dev Biol 139:325–74.
- Dial BE, Fitzpatrick LC. 1981. The energetic costs of tail autotomy to reproduction in the lizard *Coleonyx brevis* (Sauria: gekkonidae). Oecologia 51:310–7.
- Ding GH, Lin ZH, Wei L. 2014. The compensatory effect of tail regeneration on swimming speed in larval *Hoplobatrachus chinensis* Osbeck, 1765 (Anura: Ranidae) after tail removal. Acta Herpetol 9:219–25.
- Diogo R. 2020. Cranial or postcranial—dual origin of the pectoral appendage of vertebrates combining the fin-fold and gill-arch theories? Dev Dynam 249:1182–200.
- Dobrott CI, Sathyamurthy A, Levine AJ. 2019. Decoding cell type diversity within the spinal cord. Curr Opin Physiol 8:1–6.
- Donoghue PCJ, Keating JN. 2014. Early vertebrate evolution. Front Paleontol 57:875–93.
- Drucker EG, Jensen JS. 1996. Pectoral fin locomotion in the striped surfperch: i. Kinematic effects of swimming speed and body size. J Exp Biol 199:2235–42.

- Duboule D. 1998. Vertebrate *Hox* gene regulation: clustering and/or colinearity? Curr Biol 8:514–8.
- Economides KD, Zeltser L, Capecchi MR. 2003. Hoxb13 mutations cause overgrowth of caudal spinal cord and tail vertebrae. Dev Biol 256:317–30.
- Esposito CJ, Tangorra JL, Flammang BE, Lauder GV. 2012. A robotic fish caudal fin: effects of stiffness and motor program on locomotor performance. J Exp Biol 215:56–67.
- Elliot SA, Alvarado AS. 2012. The history and enduring contributions of planarians to the study of animal regeneration. Wiley Interdiscip Rev Dev Biol 2:301–26.
- El Shahawy M, Reibring C, Hallberg K, Neben CL, Marangoni P, Harfe BD, Klein OD, Linde A, Gritli-Linde A. 2019. Sonic hedgehog signaling is required for cyp26 expression during embryonic development. Int J Mol Sci 20:2275.
- Faber DS, Fetcho JR, Korn H. 1989. Neuronal networks underlying the escape response in goldfish. Ann NY Acad Sci 563:11–33.
- Felice RN, O'Connor PM. 2014. Ecology and caudal skeletal morphology in birds: the convergent evolution of pygostyle shape in underwater foraging taxa. PLoS ONE 9:e89737.
- Fish FE. 1979. Thermoregulation in the muskrat (*Ondatra zibethicus*): the use of regional heterothermia. Comp Biochem Physiol 64:391–7.
- Fish FE. 1982. Function of the compressed tail of surface swimming muskrats (*Ondatra zibethicus*). J Mammal 63:591–7.
- Fish FE. 2020. Advantages of aquatic animals as models for bio-inspired drones over present AUV technology. Bioinspir Biomim 15:025001.
- Flammang BE. 2014. The fish tail as a derivation from axial musculoskeletal anatomy: an integrative analysis of functional morphology. Zoology 117:86–92.
- Flammang BE, Tangorra JL, Mignano AP, Lauder GV. 2017. Building a fish: the biology and engineering behind an autonomous underwater vehicle. MTS J 51:15–22.
- Flammang BE, Porter ME. 2011. Bioinspiration: applying mechanical design to experimental biology. Integr Comp Biol 51:128–32.
- Fodor ACA, Powers MM, Andrykovich K, Liu J, Lowe EK, Brown CT, DiGregorio A, Stolfi A, Swalla BJ. 2021. The degenerate tale of ascidian tails. Integr Compar Biol (doi: 10.1093/icb/icab022).
- Fox SF, Conder JM, Smith AE. 1998. Sexual dimorphism in the ease of tail autotomy: Uta stansburiana with and without previous tail loss. Copeia 1998:376–82.
- Freitas R, Gómez-Marín C, Wilson JM, Casares F, Gómez-Skarmeta JL. 2012. Hoxd13 contribution to the evolution of vertebrate appendages. Dev Cell 23:1219–29.
- Freymiller GA, Whitford MD, Higham TE, Clark RW. 2019. Escape dynamics of free-ranging desert kangaroo rats (Rodentia: Heteromyidae) evading rattlesnake strikes. Biol J Linn Soc 127:164–72.
- Full RJ, Koditschek DE. 1999. Templates and anchors: neuromechanical hypotheses of legged locomotion on land. J Exp Biol 202:3325–32.
- Gardiner JD, Dimitriadis G, Codd JR, Nudds RL. 2011. A potential role for bat tail membranes in flight control. PLoS ONE 6:e18214.
- Gatesy SM, Dial KP. 1996. Locomotor modules and the evolution of avian flight. Evolution 50:331-40.

- Gehrke AR, Shubin NH. 2016. Cis-regulatory programs in the development and evolution of vertebrate paired appendages. Semin Cell Dev Biol 57:31–9.
- Geyer H, Seyfarth A, Blickhan R. 2002. Natural dynamics of spring-like running: emergence of selfstability. Paris, France: CLAWAR.
- Gibb AC, Ashley-Ross MA, Hsieh ST. 2013. Thrash, flip, or jump: the behavioral and functional continuum of terrestrial locomotion in teleost fishes. Integr Comp Biol 53:295–306.
- Gilbert EA, Payne SL, Vickaryous MK. 2013. The anatomy and histology of caudal autotomy and regeneration in lizards. Physiol Biochem Zool 86:631–44.
- Gillis GB, Bonvini LA, Irschick DJ. 2009. Losing stability: tail loss and jumping in the arboreal lizard *Anolis carolinensis*. J Exp Biol 212:604–9.
- Gillis GB, Kuo CY, Irschick DJ. 2013. The impact of tail loss on stability during jumping in green anoles (*Anolis carolinensis*). Physiol Biochem Zool 86:680–9.
- Gordeev DA, Ananjeva NB, Korost DV. 2020. Autotomy and regeneration in squamate reptiles (squamata, reptilia): Defensive behavior strategies and morphological characteristics (using computer microtomography methods). Biology Bullitin 47:389–98.
- Goulding M. 2009. Circuits controlling vertebrate locomotion: moving in a new direction. Nat Rev Neurosci 10:507–18.
- Grillner S, Wallen P, Brodin L, Lansner A. 1991. Neuronal network generating locomotor behavior in lamprey: circuitry, transmitters, membrane properties, and simulation. Ann Rev Neurosci 14:169–99.
- Hadzhiev Y, Lele Z, Schindler S, Wilson SW, Ahlberg P, Strähle U, Müller F. 2007. Hedgehog signaling patterns the outgrowth of unpaired skeletal appendages in zebrafish. BMC Dev Biol 7:75.
- Hale ME. 1996. Functional morphology of ventral tail bending and prehensile abilities of the seahorse, *Hippocampus kuda*. J Morphol 227:51–65.
- Hale ME, Long JH, McHenry MJ, Westneat MW. 2002. Evolution of behavior and neural control of the fast-start escape response. Evolution 56:993–1007.
- Han B, Zhu K, Li F, Xiae Y, Feng J, Shi Z, Lin M, Wang J, Chen Q. 2008. A simple disc degeneration model induced by percutaneous needle puncture in the rat tail. Spine 33:1925–34.
- Handa T, Ishihara H, Ohshima H, Osada R, Tsuji H, Obata K. 1997. Effects of hydrostatic pressure on matrix synthesis and matrix metalloproteinase production in the human lumbar intervertebral disc. Spine 22:1085–91.
- Hermann BG. 1995. The mouse *Brachyury* (*T*) gene. Dev Biol 6:385–94.
- Hessel AL, Nishikawa KC. 2017. The hip-twist jump: a unique mechanism for jumping in lungless salamanders. J Herpetol 51:461–7.
- Hickman GC. 1979. The mammalian tail: a review of functions. Mammal Rev 9:143–57.
- Higham TE, Russell AP. 2010. Flip, flop and fly: modulated motor control and highly variable movement patterns of autotomized gecko tails. Biol Lett 6:70–3.
- Higham TE, Russell AP, Zani PA. 2013. Integrative biology of tail autotomy in lizards. Physiol Biochem Zool 86:603–10.
- Hilde KL, Levine AJ, Hinckley CA, Hayashi M, Montgomery JM, Gullo M, Driscoll SP, Grosschedl R, Kohwi Y, Kohwi-

Shigematsu T, et al. 2016. Satb2 is required for the development of a spinal exteroceptive microcircuit that modulates limb position. Neuron 91:763–76.

- Hildebrand M. 1995. Analysis of vertebrate structure. New York (NY): Wiley.
- Ho DM, Whitman M. 2008. TGF- β signaling is required for multiple processes during *Xenopus* tail regeneration. Dev Biol 315:203–16.
- Holland P, 2013. Evolution of homeobox genes. WIREs Dev Biol 2:31-445.
- Hsieh AH, Wagner DR, Cheng LY, Lotz JC. 2005. Dependence of mechanical behavior of the murine tail disc on regional material properties: a parametric finite element study. J Biomech Eng 127:1158–67.
- Hsieh ST. 2010. A locomotor innovation enables water-land transition in a marine fish. PLoS ONE 5:e11197.
- Hsieh ST. 2016. Tail loss and narrow surfaces decrease locomotor stability in the arboreal green anole lizard (*Anolis carolinensis*). J Exp Biol 219:364–73.
- Humphreys RK, Ruxton GD. 2019. Dropping to escape: a review of an under-appreciated antipredator defence. Biol Rev 4:575–89.
- Huston SJ, Jayaraman V. 2011. Studying sensorimotor integration in insects. Curr Opin Neurobiol 21:527–34.
- Hutchins ED, Markov GJ, Eckalbar WL, George RM, King JM, Tokuyama MA, Geiger LA, Emmert N, Ammar MJ, Allen AN, et al. 2014. Transcriptomic analysis of tail regeneration in the lizard *Anolis carolinensis* reveals activation of conserved vertebrate developmental and repair mechanisms. PLoS ONE 9:e105004.
- Iatridis JC, Mente PL, Stokes IAF, Aronsson DD, Alini M. 1999. Compression-induced changes in intervertebral disc properties in a rat tail model. Spine 24:996–1002.
- Ibrahim N, Maganuco S, Dal Sasso C, Fabbri M, Auditore M, Bindellini G, Martill DM, Zouhri S, Mattarelli DA, Unwin DM, et al. 2020. Tail-propelled aquatic locomotion in a theropod dinosaur. Nature 581:67–70.
- Ishihara H, McNally DS, Urban JPG, Hall AC. 1996. Effects of hydrostatic pressure on matrix synthesis in different regions of the intervertebral disk. J Appl Physiol 80:839–46.
- Jackson J, Elgar MA. 1993. The foraging behaviour of the Willie Wagtail Rhipidura leucophrys: why does it wag its tail? Emu Austral Ornithol 93:284–6.
- Jacyniak J, McDonald RP, Vickaryous MK. 2017. Tail regeneration and other phenomena of wound healing and tissue restoration in lizards. J Exp Biol 220:2858–69.
- Jagnandan K, Higham TE. 2017. Lateral movements of a massive tail influence gecko locomotion: an integrative study comparing tail restriction and autotomy. Sci Rep 7:10865.
- Jagnandan K, Russell AP, Higham TE. 2014. Tail autotomy and subsequent regeneration alter the mechanics of locomotion in lizards. J Exp Biol 217:3891–7.
- Jankowska E. 2001. Spinal interneuronal systems: identification, multifunctional character and reconfigurations in mammals. J Physiol 533:31–40.
- Johansson LC, Hedenström A. 2009. The vortex wake of blackcaps (*Sylvia atricapilla* L.) measured using highspeed digital particle image velocimetry (DPIV). J Exp Biol 212:3365–76.

- Johnson EH, Carter AM. 2019. Defossilization: a review of 3D printing in experimental paleontology. Front Ecol Evol 7:430.
- Joven A, Elewa A, Simon A. 2019. Model systems for regeneration: salamanders. Development 146:dev167700.
- Jusufi A, Kawano DT, Libby T, Full RJ. 2010. Righting and turning in mid-air using appendage inertia: reptile tails, analytical models and bio-inspired robots. Bioinspir Biomim 5:045001.
- Kardong KV. 2019. Vertebrates: comparative anatomy, function, evolution. New York (NY): McGraw Hill.
- Kloss C, Goniva C, Hager A, Amberger S, Pirker S. 2012. Models, algorithms and validation for opensource DEM and CFD-DEM. Prog Comput Fluid Dynam Int J 12:140–52.
- Koch SC, Acton D, Goulding M. 2018. Spinal circuits for touch, pain, and itch. Ann Rev Physiol 80:189–217.
- Koch NM, Wilcoxen TE. 2019. The effects of tail damage on tadpole development and leaping ability after metamorphosis in Cuban tree frogs (*Osteopilus septentrionalis*). Bios 89:165–73.
- Kopman V, Laut J, Acquaviva F, Rizzo A, Porfiri M. 2015. Dynamic modeling of a robotic fish propelled by a compliant tail. IEEE J Ocean Eng 40:209–21.
- Lauder GV. 2000. Function of the caudal fin during locomotion in fishes: kinematics, flow visualization, and evolutionary patterns1. Am Zool 40:101–22.
- Lauder GV, Lim J, Shelton R, Witt C, Anderson E, Tangorra JL. 2011. Robotic models for studying undulatory locomotion in fishes. Mar Technol Soc J 45:41–55.
- Lemelin P. 1995. Comparative and functional myology of the prehensile tail in new world monkeys. J Morphol 224:351–68.
- Letelier J, de la Calle-Mustienes E, Pieretti J, Naranjo S, Maeso I, Nakamura T, Pascual-Anaya J, Shubin NH, Schneider I, Martinez-Morales JR, et al. 2018. A conserved *Shh* cis-regulatory module highlights a common developmental origin of unpaired and paired fins. Nat Genet 50:504–9.
- Levine AJ, Hinckley CA, Hilde KL, Driscoll SP, Poon TH, Montgomery JM, Pfaff SL. 2014. Identification of a cellular node for motor control pathways. Nat Neurosci 17:586–93.
- Levinsson A, Luo XL, Holmberg H, Schouenborg J. 1999. Developmental tuning in a spinal nociceptive system: effects of neonatal spinalization. J Neurosci 19:10397–403.
- Li C, Umbanhowar PB, Komsuoglu H, Koditschek DE, Goldman DI. 2009. Sensitive dependence of the motion of a legged robot on granular media. Proc Natl Acad Sci USA 106:3029–34.
- Libby T, Moore TY, Chang-Siu E, Li D, Cohen DJ, Jusufi A, Full RJ. 2012. Tail-assisted pitch control in lizards, robots and dinosaurs. Nature 481:181–4.
- Liem KF, Bemis WE, Walker WF Jr, Grande L. 2001. Functional anatomy of the vertebrates: an evolutionary perspective. Philadelphia (PA): Harcourt.
- Lindsey CC. 1978. Form, function, and locomotory habits in fish. In: Hoar WS, Randall DJ, editors. Fish physiology: locomotion. Vol. 7. New York (NY): Academic Press. p. 1–100.
- Lin Y, Siddall R, Schwab F, Fukushima T, Banerjee H, Baek Y, Vogt D, Park Y, Jusufi A. 2021. Modeling and control of a soft robotic fish with integrated soft sensing. Advancing

Intelligent Systems 2000244. https://doi.org/10.1002/aisy. 202000244

- Liu Y, Ben-Tzvi P. 2021. Dynamic modeling, analysis, and design synthesis of a reduced complexity quadruped with a serpentine robotic tail. Integr Compar Biol. DOI: 10.1093/icb/icab083
- Long JH, Koob TJ, Irving K, Combie K, Engel V, Livingston N, Lammert A, Schumacher J. 2006. Biomimetic evolutionary analysis: testing the adaptive value of vertebrate tail stiffness in autonomous swimming robots. J Exp Biol 209:4732–46.
- Longellato F, Fiorentino M, De Felice E, Luigi C, Nittoli V, Joss JMP, Sordino P. 2018. Expression of *meis* and *hoxa11* in dipnoan and teleost fins provides new insights into the evolution of vertebrate appendages. Evodevo 9:11.
- Lotfollahi M, Naghipourfar M, Luecken M, Khajavi M, Büttner M, Avsec Z, Misharin A, Theis F. 2020. Query to reference single-cell integration with transfer learning. bioRxiv 205997:1–26.
- Luger AM, Ollevier A, De Kegel B, Herrel A, Adriaens D. 2020. Is variation in tail vertebral morphology linked to habitat use in chameleons? J Morphol 281:229–39.
- Lundberg A. 1979. Multisensory control of spinal reflex pathways. Prog Brain Res 50:11–28.
- Martin BL, Kimelman D. 2008. Regulation of canonical *Wnt* signaling by *Brachyury* is essential for posterior mesoderm formation. Dev Cell 15:121–33.
- Marvi H, Gong C, Gravish N, Astley H, Travers M, Hatton RL, Mendelson JR, Choset H, Hu DL, Goldman DI. 2014. Sidewinding with minimal slip: snake and robot ascent of sandy slopes. Science 346:224–49.
- Marvin GA. 2010. Effect of caudal autotomy on aquatic and terrestrial locomotor performance in two desmognathine salamander species. Copeia 3:468–74.
- Marvin GA. 2011. Effect of body size on tail regeneration and recovery of swimming performance after caudal autotomy in a plethodontid salamander. Amphibia-Reptilia 32:485–92.
- Marvin GA. 2013. Critical tail autotomy for reduction of maximal swimming performance in a plethodontid salamander (*Desmognathus quadramaculatus*). J Herpetol 47:174–8.
- Marvin GA, Lewis M. 2013. Effect of temperature, photoperiod, and feeding on the rate of tail regeneration in a semiaquatic plethodontid salamander. J Thermal Biol 38:548–52.
- Mathis A, Mamidanna P, Cury KM, Abe T, Murthy VN, Mathis MW, Bethge M. 2018. DeepLabCut: markerless pose estimation of user-defined body parts with deep learning. Nat Neurosci 21:1281–9.
- Maybury WJ, Rayner JMV, Couldrick LB. 2001. Lift generation by the avian tail. Proc R Soc Lond B Biol Sci 268:1443–8.
- Maybury WJ, Rayner JMV. 2001. The avian tail reduces body parasite drag by controlling flow separation and vortex shedding. Proc R Soc Lond B Biol Sci 268:1405–10.
- McCafferty DJ. 2007. The value of infrared thermography for research on mammals: previous applications and future directions. Mammal Rev 37:207–23.
- McHenry MJ, Feitl KE, Strother JA, Van Trump WJ. 2009. Larval zebrafish rapidly sense the water flow of a predator's strike. Biol Lett 5:477–9.

- McInroe B, Astley HC, Gong C, Kawano SM, Schiebel PE, Rieser JM, Choset H, Blob RW, Goldman DI. 2016. Tail use improves performance on soft substrates in models of early vertebrate land locomotors. Science 353:154–8.
- McMahon TA, Cheng GC. 1990. The mechanics of running: how does stiffness couple with speed? J Biomech 23:65–78.
- Mentel T, Krause A, Pabst M, El Manira A, Büschges A. 2006. Activity of fin muscles and fin motoneurons during swimming motor pattern in the lamprey. Eur J Neurosci 23:2012–26.
- Mercader N. 2007. Early steps of paired fin development in zebrafish compared with tetrapod limb development. Dev Growth Differ 49:421–37.
- Moore J, Gutmann A, McGowan CP, McKinley P. 2013. Exploring the role of the tail in bipedal hopping through computational evolution. In: Artificial Life Conference Proceedings, Cambridge, MA. p. 11–8.
- Møller A, Nielsen JT, Erritzøe J. 2006. Losing the last feather: feather loss as an antipredator adaptation in birds. Behav Ecol 17:1046–56.
- Müller UK, Smit J, Stamhuis EJ, Videler JJ. 2001. How the body contributes to the wake in undulatory fish swimming: flow fields of a swimming eel (*Anguilla anguilla*). J Exp Biol 204:2751–62.
- Nakanishi ST, Whelan PJ. 2012. A decerebrate adult mouse model for examining the sensorimotor control of locomotion. J Neurophysiol 107:500–15.
- Neutens C, Adriaens D, Christiaens J, De Kegel B, Dierick M, Boistel R, Van Hoorebeke L. 2014. Grasping convergent evolution in syngnathids: a unique tale of tails. J Anat 224:710–23.
- Nishikawa K, Biewener AA, Aerts P, Ahn AN, Chiel HJ, Daley MA, Daniel TL, Full RJ, Hale ME, Hedrick TL, et al. 2007. Neuromechanics: an integrative approach for understanding motor control. Integr Comp Biol 47:16–54.
- O'Connor SM, Dawson TJ, Kram R, Donelan JM. 2014. The kangaroo's tail propels and powers pentapedal locomotion. Biol Lett 10:20140381.
- Oliver SP, Turner JR, Gann K, Silvosa M, D'Urban Jackson T. 2013. Thresher sharks use tail-slaps as a hunting strategy. PLoS ONE 8:e67380.
- Patel A, Braae M. 2013. Rapid turning at high-speed: Inspirations from the cheetah's tail. In: 2013 IEEE/RSJ International Conference on Intelligent Robots and Systems, Tokyo, Japan. p. 5506–11.
- Patel A, Boje E. 2015. On the conical motion of a two-degreeof-freedom tail inspired by the cheetah. IEEE Trans Robot 31:1555–60.
- Patel A, Boje E, Fisher C, Louis L, Lane E. 2016. Quasisteady state aerodynamics of the cheetah tail. Biol Open 5:1072–6.
- Peixoto LT, Kolmann MA, Pfeiffenberger J, Summers AP, Donatelli CM. 2019. Not so ridged after all: ontogeny of armor biomechanics and swimming performance in poachers. J Morphol 280:168–9.
- Pianka ER. 1973. The structure of lizard communities. Ann Rev Ecol Syst 4:53–74.
- Picton LD, Bertuzzi M, Pallucchi I, Fontanel P, Dahlberg E, Björnfors ER, Iacoviello F, Shearing PR, El Manira A. 2021. A spinal organ of proprioception for integrated motor action feedback. Neuron 109:1–14.

- Plimpton S. 1995. Fast parallel algorithms for short-range molecular dynamics. J Comput Phys 117:1–19.
- Porter MM, Adriaens D, Hatton RL, Meyers MA, McKittrick J. 2015. Why the seahorse tail is square. Science 349:aaa6683.
- Rashid DJ, Surya K, Chiappe LM, Carroll N, Garrett KL, Varghese B, Bailleul A, O'Connor JK, Chapman SC, Horner JR. 2018. Avian tail ontogeny, pygostyle formation, and interpretation of juvenile Mesozoic specimens. Sci Rep 8:9014.
- Reeves WM, Shimai K, Winkley KM, Veeman MT. 2021. *Brachyury* controls *Ciona* notochord fate as part of a feedforward network and not as a unitary master regulator. Development 148:dev195230.
- Rone WS, Ben-Tzvi P. 2014. Continuum robotic ail loading analysis for mobile robot stabilization and maneuvering. *Proceedings of the ASME 2014 International Design Engineering Technical Conferences.* Buffalo, New York, USA.
- Rone W, Ben-Tzvi P. 2015. Dynamic modeling and simulation of a yaw-angles quadruped maneuvering with a planar robotic tail. J Dyn Syst Meas Control 138:084502.
- Rosic MN, Thornycroft PJM, Feilich KL, Lucas KN, Lauder GV. 2017. Performance variation due to stiffness in a tunainspired flexible foil model. Bioinspir Biomim 12:016011.
- Saab W, Rone WS, Ben-Tzvi P. 2018. Robotic tails: a state-ofthe-art review. Robotic. DOI: 10.1017/s0263574718000425.
- Sachs G. 2007. Tail effects on yaw stability in birds. J Theor Biol 249:464–72.
- Sallan L. 2016. Fish 'tails' result from outgrowth and reduction of two separate ancestral tails. Curr Biol 26:R1224-5.
- Salvador A, Martin J, Lopez P. 1995. Tail loss reduces home range size and access to females in male lizards *Psammodromus algirus*. Behav Ecol 6:382–7.
- Schmitt J, Holmes P. 2000. Mechanical models for insect locomotion: dynamics and stability in the horizontal plane-II. Appl Biol Cybern 83:517–27.
- Schouenborg J. 2008. Action-based sensory encoding in spinal sensorimotor circuits. Brain Res Rev 57:111–7.
- Schouenborg J, Weng HR. 1994. Sensorimotor transformation in a spinal motor system. Exp Brain Res 100:170–4.
- Schouenborg J, Weng HR, Kalliomaki J. 1995. A survey of spinal doral horn neurons encoding the spatial organization of withdrawal reflexes in the rat. Exp Brain Res 106:19–27.
- Schulte-Merker S. 1995. The zebrafish no tail gene. Dev Biol 6:411-6.
- Schuster S. 2012. Fast-starts in hunting fish: decision-making in small networks of identified neurons. Curr Opin Neurobiol 22:279–84.
- Schwaner MJ, Freymiller GA, Clark RW, McGowan CP. 2021. How to stick the landing: kangaroo rats use their tails to reorient during evasive jumps away from predators. Integr Comp Biol. DOI: 10.1093/icb/icab043
- Seyfarth A, Geyer H, Herr H. 2003. Swing-leg retraction: a simple control model for stable running. J Exp Biol 206:2547–455.
- Shen ZH, Seipel JE. 2012. A fundamental mechanism of legged locomotion with hip torque and leg damping. Bioinspir Biomim 7:046010.

- Spence AJ, Revzen S, Seipel J, Mullens C, Full RJ. 2010. Insects running on elastic surfaces. J Exp Biol 213:1907–20.
- Sherrington CS. 1908. The integrative action of the nervous system. Science 27:885–9.
- Sherrington CS. 1913. Further observations on the production of reflex stepping by combination of reflex excitation with reflex inhibition. J Physiol 47:196–214.
- Shield S, Jericevich R, Patel A, Jusufi A. 2021. Tails, flails, and sails: how appendages improve terrestrial maneuverability by improving stability. Integr Comp Biol.
- Shubin N, Tabin C, Carroll S. 1997. Fossils, genes and the evolution of animal limbs. Nature 388:639–48.
- Sleight VA, Gillis JA. 2020. Embryonic origin and serial homology of gill arches and paired fins in the skate, *Leucoraja erinacea*. eLife 9:e60635.
- Skeels A, Esquerré D, Cardillo M. 2020. Alternative pathways to diversity across ecologically distinct lizard radiations. Glob Ecol Biogeogr 29:454–69.
- Spenko MJ, Haynes GC, Saunders JA, Cutkosky MR, Rizzi AA, Full RJ, Koditschek DE. 2008. Biologically inspired climbing with a hexapedal robot. J Field Robot 25:223–42.
- Swanson BO, Gibb AC. 2004. Kinematics of aquatic and terrestrial escape responses in mudskippers. J Exp Biol 207:4037–44.
- Thisse B, Pflumio S, Fürthauer M, Loppin B, Heyer V, Degrave A, Woehl R, Lux A, Staffan T, Charbonnier XQ, Thisse C. 2001. Expression of the zebrafish genome during embryogenesis (NIH R01 RR15402). ZFIN Direct Data Submission. (http://zfin.org).
- Thomas ALR. 1997. On the tails of birds. Bioscience 47:215–25.
- Triantafyllou MS, Triantafyllou GS. 1995. An efficient swimming machine. Sci Am 272:64–71.
- Triay-Portella R, Gonzalez JA, Pajuelo JG. 2019. Caudal region regeneration in a natural population of *Physiculus cyanostrophus* in the tropical eastern Atlantic ocean. Deep-Sea Res I 150:103062.
- Tucker VA. 1992. Pitching equilibrium, wing span and tail span in a gliding Harris' Hawk, *Parabuteo unicinctus*. J Exp Biol 165:21–41.
- Tulenko FJ, Augustus GJ, Massey JL, Sims SE, Mazan S, Davis MC. 2016. *HoxD* expression in the fin-fold compartment of basal gnathostomes and implications for paired appendage evolution. Sci Rep 6:22720.
- Tytell ED, Lauder GV. 2002. The C-start escape response of *Polypterus senegalus*: bilateral muscle activity and variation during stage 1 and 2. J Exp Biol 205:2591–603.
- Tytell ED, Lauder GV. 2004. The hydrodynamics of eel swimming: i. Wake structure. J Exp Biol 207:1825–41.
- Uemura Y, Kato K, Kawakami K, Kimura Y, Oda Y, Higashijima SI. 2020. Neuronal circuits that control rhythmic pectoral fin movements in Zebrafish. J Neurosci 40:6678–90.
- Usherwood JR, Cheney JA, Song J, Windsor SP, Stevenson JPJ, Dierksheide U, Nila A, Bomphrey RJ. 2020. High aerodynamic lift from the tail reduces drag in gliding raptors. J Exp Biol 223:jeb214809.
- Verissimo KM, Perez NL, Dragalzew AC, Senevirathne G, Darnet S, Mendes WRB, Dos Santoos Neves CA, Dos Santos EM, de Sousa Moraes CN, Elewa A, et al. 2020.

Salamander-like tail regeneration in the West African lungfish. Proc Biol Soc 287:20192939.

- Vitt LJ, Congdon JD, Dickson NA. 1977. Adaptive strategies and energetics of tail autonomy in lizards. Ecology 58:326–37.
- Vitulo N, Valle LD, Skobo T, Valle G, Alibardi L. 2016. Transcriptome analysis of the regenerating tail vs. the scarring limbs in lizards reveals pathways leading to successful vs. unsuccessful organ regeneration in amniotes. Dev Dyn 246:116–34.
- Wake DB, Dresner IG. 1967. Functional morphology and evolution of tail autotomy in salamanders. J Morphol 122:265–305.
- Watkins-Colwell G, Love K, Randall Z, Boyer D, Winchester J, Stanley E, Blackburn D. 2018. The walking dead: status report, data workflow and best practices of the overt thematic collections network. Biodivers Inform Sci Stand 2:e26078.
- Webb PW. 2002. Control of posture, depth, and swimming trajectories of fishes. Integr Comp Biol 42:94–101.
- Webster FA, Griffin DR. 1962. The role of the flight membranes in insect capture by bats. Anim Behav 10:332–40.
- Wilson AM, Lowe JC, Roskilly K, Hudson PE, Golabek KA, McNutt JW. 2013. Locomotion dynamics of hunting in wild cheetahs. Nature 498:185–9.
- Wilson V, Rashbass P, Beddington RSP. 1993. Chimeric analysis of *T* (*Brachyury*) gene function. Development 117:1321–31.
- Woodland DJ, Jaafar Z, Knight ML. 1980. The "pursuit deterrent" function of alarm signals. Am Nat 115:748–53.
- Wyart C, Knafo S. 2015. Sensorimotor integration in the spinal cord, from behaviors to circuits: new tools to close the

loop? In: Douglass AD, editor. New techniques in systems neuroscience. Berlin/Heidelberg, Germany: Springer. p. 197–234.

- Xiong Z, Li F, Li Q, Zhou L, Gamble T, Zheng J, Kui L, Li C, Li S, Yang H, et al. 2016. Draft genome of the leopard gecko *Eublepharis macularius*. GigaScience 5.
- Xu C, Hutchins ED, Tokuyama MA, Rawls JW, Kusumi K. 2020. Transcriptional analysis of scar-free wound healing during early stages of tail regeneration in the green lizard, *anolis carolinensis*. J Immunol Regen Med 7:100025.
- Xu C, Palade J, Fisher RE, Smith CI, Clark AR, Sampson S, Bourgeois R, Rawls A, Elsey RM, Wilson-Rawls J, et al. 2020. Anatomical and histological analyses reveal that tail repair is coupled with regrowth in wild-caught, juvenile American alligators (*Alligator mississippiensis*). Sci Rep 10:20122.
- Yano T, Tamura K. 2013. The making of differences between fins and limbs. J Anat 222:100–13.
- Yurewicz KL, Wilbur HM. 2004. Resource availability and costs of reproduction in the salamander *Plethodon cinereus*. Copeia 1:28–36.
- Zamora-Camacho FJ, Cortés-Manzaneque S, Aragón P. 2019. Simulated predation pressure in Pelobates cultripes tadpoles modulates morphology at the metamorphic stage. Curr Zool 65:651–6.
- Zhang X, Gong J, Yao Y. 2016. Effects of head and tail as swinging appendages on the dynamic walking performance of a quadruped robot. Robotica 34:2878–91.
- Zhu J, White C, Wainwright DK, Di Santo V, Lauder GV, Bart-Smith H. 2019. Tuna robotics: a high-frequency experimental platform exploring the performance space of swimming fishes. Sci Robot 4:eaax4615.